

**CHROMOSOME POLYMORPHISM IN *CHIRONOMUS
BALATONICUS* DEVAI, WÜLKER, SCHOLL (DIPTERA,
CHIRONOMIDAE) FROM BOURGAS LAKE**

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ABSTRACT. Seasonal and year fluctuations of chromosome polymorphism in *C. balatonicus* from Bourgas Lake were investigated. The chromosome polymorphism features – frequency of larvae with heterozygous inversions and heterozygous inversions per larva showed seasonal dependence: increased in spring and summer and decreased in winter. Statistically significant correlation between seasonal fluctuations of the temperature and frequency of specimens with the heterozygous inversion A12 was also established.

Besides structural chromosome rearrangements high functional activity was found: appearance of new puffs, ectopic pairing between the chromosomes, decondensation of the telomeres and heterozygous state in BR₂. These structural and functional chromosome aberrations in the light of the impact of abiotic factors on the chironomids karyotype are discussed.

KEY WORDS. Chironomidae, *Chironomus balatonicus*, polytene chromosomes, chromosome aberrations

INTRODUCTION

Chironomus balatonicus DEVAI, WÜLKER, SCHOLL, 1983, is a widely distributed species in Palearctic. This species is characterized by high level of chromosome and genome polymorphism (KIKNADZE *et al.* 1991, BELJANINA, LOGINOVA 1993, MICHAILOVA 1989, 2001, MICHAILOVA, PETROVA 1994, MICHAILOVA, KRASTANOV 2000, MICHAILOVA *et al.* 2002). The chromosome polymorphism is realized by heterozygous and homozygous inversions. Some of these inversions form polymorphous system characterized by transition of one homozygous inversion into another through heterozygotization (MICHAILOVA, PETROVA 1994, PETROVA *et al.* 1994).

The genome polymorphism is expressed with an additional "B" chromosome (PETROVA 1991, MICHAILOVA, PETROVA 1994, GOLYGINA *et al.* 1996).

The purpose of the present study is to clarify chromosomal polymorphism and seasonal and year fluctuation in chromosome variability in *Chironomus balatonicus* from Bourgas Lake.

MATERIAL AND METHODS

The larval material has been collected in the mud from the bottom of Bourgas Lake during the period X.2001-X.2003. The larvae were fixed in a 3:1 mixture of 96% ethanol and glacial acetic acid. Polytene chromosome preparations were prepared by routine aceto-orcein method (MICHAILOVA 1989). The cytotaxonomical differentiation of the larval material has been done according to Michailova (1989) and Kiknadze *et al.* (1991).

The frequency of aberrations was calculated in percentage. Deviation of heterozygous inversions by Hardy-Weinberg equilibrium was statistically analyzed using χ^2 method. The statistical significance of the heterozygotes frequency between the samples by the different years was tested with G-test. Correlation analysis was used to establish dependence between variation of the chromosome aberrations and the seasonal fluctuation of some abiotic factors. Software programs package "Easy-Stat", version 2.3 (1991) both for "G-test" and "Correlation analysis" were used.

The chemical analysis of superficial water of Bourgas Lake was provided from the Regional Agency of Nature and Water Protection – Bourgas.

RESULTS

Chironomus balatonicus from Bourgas lake shows the same karyological characteristics as the other Palaearctic populations (DEVAI *et al.* 1983, KIKNADZE *et al.* 1991, MICHAILOVA *et al.* 2002.). This species belong to *thummi* complex and has $2n = 8$ chromosomes. Chromosomes AB and CD are metacentric, chromosome EF – submetacentric and chromosome G- telocentric. One Balbiani ring BR₁ is localized of chromosome AB (arm B). Chromosome G carries two Balbiani rings (BR₂ and BR₃) and a nucleolus (NOR).

Chironomus balatonicus is the only species identified cytotaxonomically in Bourgas lake.

In 98% of studied specimens heterozygous inversions have been established. The average number of heterozygous inversions per larva was 1,5.

In arms B, D and G, the standard banding pattern predominated. In arms A and C, the heterozygotes (A12, C12) were most frequent. In chromosome AB, heterozygous pericentric inversion has been found in 20% of the specimens. The arms E and F were monomorphous. The frequency of heterozygous inversion A12 showed statistically significant values both in 2002 and 2003 year. In 2002 it was more frequent while in 2003 was considerably lower. Significant dependence between seasonal variation of the temperature and frequency of individuals with the same inversion (A12) was established ($P < 0.05$), (Fig. 1). The observed heterozygotes and heterozygous inversion per larvae showed also seasonal dependence increased in

spring and summer and decreased in winter ($P < 0.05$), (Fig. 2). Annual fluctuations in frequency of heterozygous inversions in arms A and G were also found (Fig. 3). The frequency of the inversion A12 in 2002, was significantly different from those in 2001 and in 2003 years ($P < 0.05$, $P < 0.001$), while inversion G12, in 2003 year differed from those in 2001 and in 2002 years ($P < 0.05$).

High functional activity of the polytene chromosomes was found. In 7% of individuals an active puff near the centromere in arms A and B respectively were detected. In all individuals with the inversion D13 an active puff in the homologue D3 of arm D was also observed. All chromosome arms were involved in ectopic contacts with almost equal frequency (18-20%). With low frequency (13%) in summer larvae only decondensation of the telomeres of all chromosome arms was observed. In 27% of studied larvae heterozygous BR3 was found.

DISCUSSION

The study of seasonal dynamic of the chromosome polymorphism in *Chironomus balatonicus* from Bourgas Lake showed statistically significant coincidence in the fluctuation of the inversion heterozygotes A12 and seasonal variations of the temperature. Such a dependency was established in *Chironomus balatonicus* for the first time. In the sibling species of *Chironomus balatonicus* – *C. plumosus* LINNAEUS, 1758, connection between frequencies of the heterozygotes B12 and the depth, in which the larvae live, is established. Larvae living in the greatest depth display the highest frequencies of these heterozygotes (PEDESEN 1978). In this species from Novgorod the same inversion is considerably increased in the spring and decreased in autumn and winter (ILYNSKAJA *et al.* 1999). The peak of heterozygosity has been observed in summer and a minimum in spring and autumn. Similar dependency was detected in *C. balatonicus* from Bourgas Lake: the frequency of all larvae with heterozygous inversions and heterozygous inversions per larva were high in spring and summer and lower in winter.

In 2002 the frequency of heterozygous larvae, was higher than those in 2001 and 2003. In this year the frequency of heterozygous inversion A12 was also significantly high and considerably deviated from Hardy-Weinberg equilibrium. It is very important to note that in this year the chemical oxygen demand (COD), exceed the limit of tolerance. The high organic pollution in this year is probably the reason for the observed deviation. High frequency of structural and functional chromosome alterations in stress ecological conditions was also established in other species from genus *Chironomus*. In *C. riparius* MEIGEN, 1804, from regions with increased amounts of heavy metals, are registered different somatic structural aberrations and functional alteration (inductions of novel puffs, telomeric and centromeric decondensations) (MICHAILOVA *et al.* 2001). Ectopic pairing between the chromosomes and heterozygous state of BR₂ were observed in *Chironomus balatonicus* from Chernobil (MICHAILOVA, PETROVA 1994) and from Bulgaria in heavy metal polluted biotopes (ILKOVA 2004). The genome reactions of *Chironomus balatonicus* from Bourgas Lake, where the pH, organic and inorganic pollution were upper the standard value are very similar: new puffs, ectopic

conjugation between the chromosomes, change of the heterohromatin appearance in the telomeres regions and heterozygous state in BR₂.

It might be possible the anthropogenic pollution in the lake, significantly exceeded the values accepted as standard is the reason for such high structural and functional activity in *Chironomus balatonicus*, but also complex impact both of anthropogenic and abiotic factors of the environment are possible.

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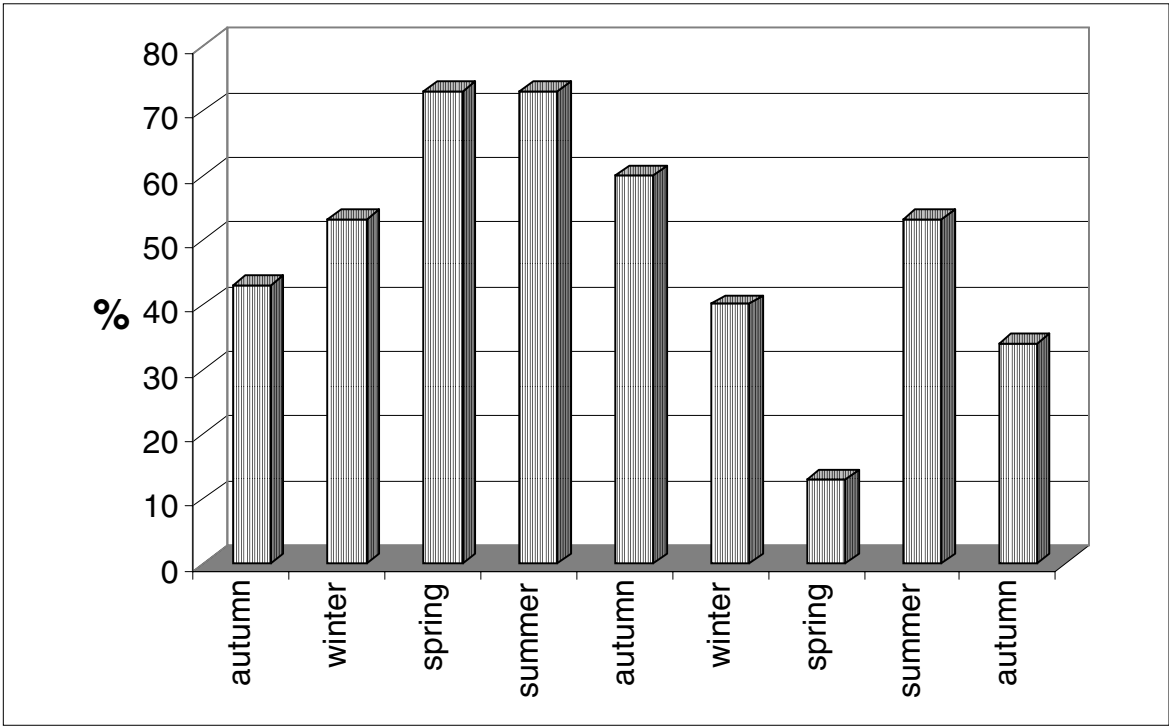


Figure 1. Seasonal fluctuation in frequency of specimens with the heterozygous inversion A12.

Denotes statistically significant values in frequency of heterozygous inversion A12 in spring both in 2002 and 2003 year.

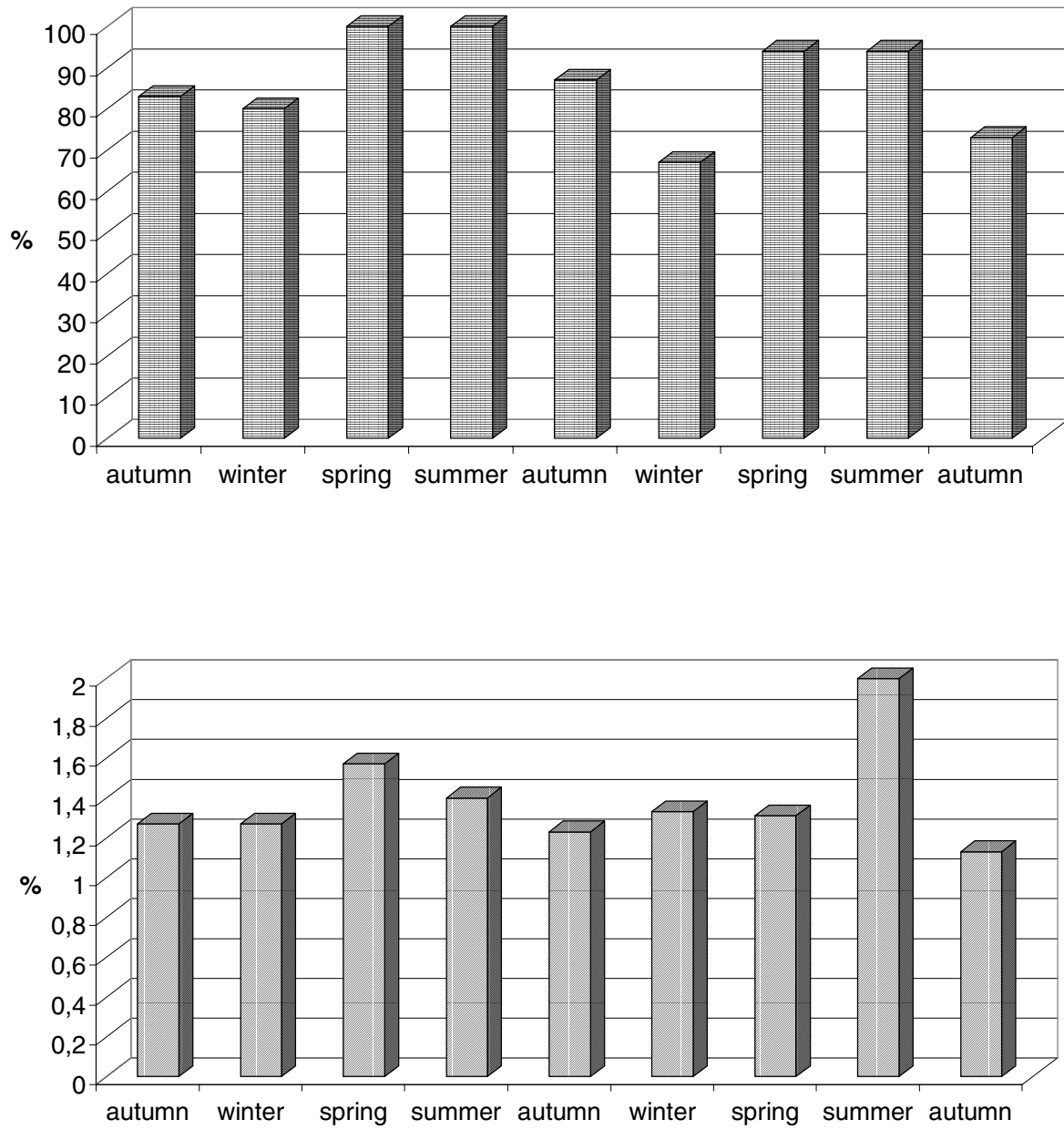


Figure 2. Seasonal alteration in the chromosome polymorphism features:
a) Frequency of larvae with heterozygous inversions.
b) Heterozygous inversion per larva.

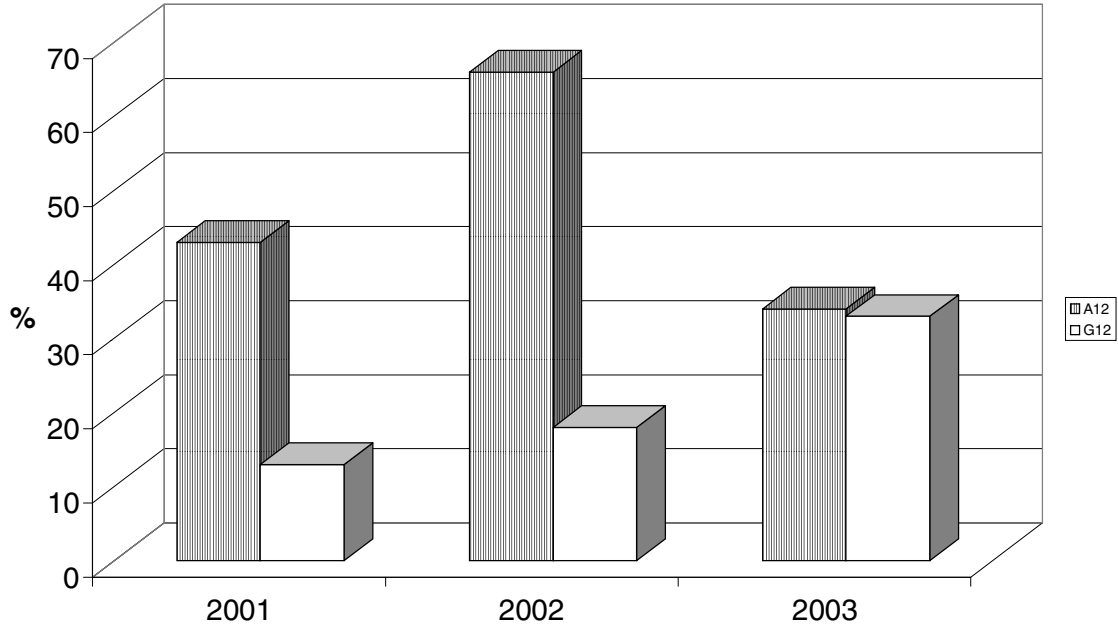


Figure 3. Annual variation in frequency of heterozygous inversions in the arms A and G.